

INSECTICIDAL PROTEINS FROM *BACILLUS THURINGIENSIS*

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INTRODUCTION

The present invention relates to new DNA sequences encoding insecticidal proteins produced by *Bacillus thuringiensis* strains. Particularly, new DNA sequences encoding proteins designated as Cry9Fa, Cry1Jd, and Cry1Bf are provided which are useful to protect plants from insect damage. Also included herein are micro-organisms and plants transformed with at least one of the newly isolated genes so that they are useful to confer insect resistance by expression of insecticidal protein.

BACKGROUND OF THE INVENTION

(i) Field of the Invention:

Bt or *Bacillus thuringiensis* is well known for its specific toxicity to insect pests, and has been used since almost a century to control insect pests of plants. In more recent years, transgenic plants expressing *Bt* proteins were made which were found to successfully control insect damage on plants (e.g., Vaeck et al., 1987).

Despite the isolation of a number of *Bt* crystal protein genes, the search for new genes encoding insecticidal proteins continues. Indeed, insecticidal *Bt* crystal proteins are known to have a relatively narrow target insect range compared to chemical insecticides. Also, having multiple toxins active on the same target insect species allows the use of proteins having different modes of action so that insect resistance development can be prevented or delayed.

(ii) Description of Related Art:

Previously, several types of Cry1B-, Cry1J-, and Cry9-proteins were identified (see Crickmore et al., 1998, incorporated herein by reference, for all details).

The new Cry1Bf protein has the closest sequence identity with the Cry1Be protein (Payne et al, 1998, US Patent 5,723,758), but still differs in about 14 percent of the amino acid sequence of its toxic protein fragment with the toxic fragment of the Cry1Be protein.

The closest sequence identity with the Cry1Jd toxic fragment was found in the toxic fragment of the Cry1Jc1 protein (US Patent 5,723,758), but the toxic fragments of both proteins still differ in about 18 % of their amino acid sequence.

The closest sequence identity with the Cry9Fa toxic fragment was found with the toxic fragment of the Cry9Ea1 protein as described by Midoh et al. (PCT Patent publication WO 98/26073) and Narva et al. (PCT patent publication WO 98/00546), but the toxic fragments of the Cry9Fa and Cry9Ea proteins still differ in about 21 % of their amino acid sequence.

SUMMARY OF THE INVENTION

In accordance with this invention is provided a DNA sequence encoding a protein comprising the amino acid sequence selected from the group consisting of: a) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry1Bf* gene deposited at the BCCM-LMBP under accession number LMBP 3986, b) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry1Jd* gene deposited at the BCCM-LMBP under accession number LMBP 3983, and c) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry9Fa* gene deposited at the BCCM-LMBP under accession number LMBP 3984.

Particularly preferred in accordance with this invention is a DNA sequence encoding a protein comprising the amino acid sequence selected from the group consisting of: the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 2, the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 4, and the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 6; alternatively, a DNA encoding a protein comprising the amino acid sequence of the group selected from: the amino acid sequence of SEQ ID No. 2, the amino acid sequence of SEQ ID No. 4, the amino acid sequence of SEQ ID No. 6; or a DNA sequence comprising the DNA sequence of SEQ ID No. 1, SEQ ID No. 3, or SEQ ID No. 5..

Further, in accordance with this invention are provided DNA sequences encoding at least the following portions of the newly-isolated proteins: the amino acid sequence of SEQ ID No. 2 from amino acid position 1 to amino acid position 640, the amino acid sequence of SEQ ID No. 4 from amino acid position 1 to amino acid position 596, and the amino acid sequence of SEQ ID No. 6 from amino acid position 1 to amino acid position 652.

Further, in accordance with this invention are provided the above DNA sequences comprising an artificial DNA sequence having a different codon usage compared to the naturally occurring DNA sequence but encoding the same protein or its insecticidal fragment.

Even further provided in accordance with this invention is a protein comprising the amino acid sequence selected from the group consisting of: a) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry1Bf* gene deposited at the BCCM-LMBP under accession number LMBP 3986, b) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry1Jd* gene deposited at the BCCM-LMBP under accession number LMBP 3983, and c) the amino acid sequence of the insecticidal trypsin-digestion fragment of the protein encoded by the *cry9Fa* gene deposited at the BCCM-LMBP under accession number LMBP 3984.

Particularly preferred herein is a protein comprising the amino acid sequence selected from the group consisting of: the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 2, the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 4, and the amino acid sequence of an insecticidal fragment of the protein of SEQ ID No. 6; alternatively a protein, comprising the amino acid sequence selected from the group consisting of: the amino acid sequence of SEQ ID No. 2 from amino acid position 1 to amino acid position 640, the amino acid sequence of SEQ ID No. 4 from amino acid position 1 to amino acid position 596, and the amino acid sequence of SEQ ID No. 6 from amino acid position 1 to amino acid position 652; or a protein comprising the amino acid sequence of SEQ ID No. 2, SEQ ID No. 4, or SEQ ID No. 6.

Also provided herein are chimeric genes comprising the DNA as defined above under the control of a plant-expressible promoter, and plant cells, plants or seeds transformed to contain those chimeric genes, particularly plant cells, plants, or seeds selected from the group consisting of: corn, cotton, rice, oilseed rape, Brassica species, eggplant, soybean, potato, sunflower, tomato, sugarcane, tea, beans, tobacco, strawberry, clover, cucumber, watermelon, pepper, oat, barley, wheat, dahlia, gladiolus, chrysanthemum, sugarbeet, sorghum, alfalfa, and peanut. In accordance with this invention, the chimeric gene can be integrated in the nuclear or chloroplast DNA of the plant cells.

Further in accordance with this invention are provided micro-organisms, transformed to contain any of the above DNA sequences, particularly those selected from the genus *Agrobacterium*, *Escherichia*, or *Bacillus*.

Also provided herein is a process for controlling insects, comprising expressing any of the above DNA sequences in a host cell, particularly plant cells, and contacting insects with said host cells, and a process for rendering a plant resistant to insects, comprising transforming plants cells with any of the above DNA sequences or chimeric genes, and regenerating transformed plants from such cells which are resistant to insects.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

In accordance with this invention, DNA sequences encoding new *Bt* toxins have been isolated and characterized. The new genes were designated *cry1Bf*, *cry1Jd* and *cry9Fa*, and their encoded proteins Cry1Bf, Cry1Jd and Cry9Fa.

In accordance with this invention "Cry1Bf protein" refers to any protein comprising the smallest protein fragment of the amino acid sequence of SEQ ID No. 2 which retains insecticidal activity, particularly any protein comprising the amino acid sequence from the amino acid at position 1 to the amino acid at position 640 in SEQ ID No. 2, including but not limited to the complete protein with the amino acid sequence of SEQ ID No. 2. This includes

hybrids or chimeric proteins comprising the smallest toxic protein fragment, as well as proteins containing at least one of the three functional domains of the toxic fragment of SEQ ID No. 2. The term "DNA/protein comprising the sequence X", as used herein, refers to a DNA or protein including or containing at least the sequence X, so that other nucleotide or amino acid sequences can be included at the 5' (or N-terminal) and/or 3' (or C-terminal) end, e.g. (the nucleotide sequence of) a selectable marker protein as disclosed in EP 0 193 259.

In accordance with this invention, "Cry9Fa protein" or "Cry9F protein" refers to any protein comprising the smallest protein fragment of the amino acid sequence of SEQ ID No. 6 which retains insecticidal activity, particularly any protein comprising the amino acid sequence from the amino acid at position 1 to the amino acid at position 652 in SEQ ID No. 6, including but not limited to the complete protein with the amino acid sequence of SEQ ID No. 6. This includes hybrids or chimeric proteins comprising the smallest toxic protein fragment, as well as proteins containing at least one of the three functional domains of the toxic fragment of SEQ ID No. 6.

In accordance with this invention, "Cry1Jd protein" refers to any protein comprising the smallest protein fragment of the amino acid sequence of SEQ ID No. 4 which retains insecticidal activity, particularly any protein comprising the amino acid sequence from the amino acid at position 1 to the amino acid at position 596 in SEQ ID No. 4, including but not limited to the complete protein with the amino acid sequence of SEQ ID No. 4. This includes hybrids or chimeric proteins comprising the smallest toxic protein fragment, as well as proteins containing at least one of the three functional domains of the toxic fragment of SEQ ID No. 4.

As used herein, the terms "*cry1Bf* DNA", "*cry9Fa* DNA", or "*cry1Jd* DNA", refer to any DNA sequence encoding the Cry1Bf, Cry9Fa, or Cry1Jd protein, respectively, as defined above. This includes naturally occurring, artificial or synthetic DNA sequences encoding the newly isolated proteins or their insecticidal fragments as defined above. Also included herein are DNA sequences encoding insecticidal proteins which are similar enough to the coding regions of the genomic DNA sequences deposited or the sequences provided in the sequence listing so that they can (i.e., have the ability to) hybridize to these DNA sequences under stringent

hybridization conditions. Stringent hybridization conditions, as used herein, refers particularly to the following conditions: immobilizing the relevant genomic DNA sequences on a filter, and prehybridizing the filters for either 1 to 2 hours in 50 % formamide, 5 % SSPE, 2x Denhardt's reagent and 0.1 % SDS at 42 ° C or 1 to 2 hours in 6x SSC, 2xDenhardt's reagent and 0.1 % SDS at 68 °C. The denatured labeled probe is then added directly to the prehybridization fluid and incubation is carried out for 16 to 24 hours at the appropriate temperature mentioned above. After incubation, the filters are then washed for 20 minutes at room temperature in 1x SSC, 0.1 % SDS, followed by three washes of 20 minutes each at 68 °C in 0.2 x SSC and 0.1 % SDS. An autoradiograph is established by exposing the filters for 24 to 48 hours to X-ray film (Kodak XAR-2 or equivalent) at -70 °C with an intensifying screen. Of course, equivalent conditions and parameters can be used in this process while still retaining the desired stringent hybridization conditions. One of such equivalent conditions includes: immobilizing the relevant genomic DNA sequences on a filter, and prehybridizing the filters for either 1 to 2 hours in 50 % formamide, 5 % SSPE, 2x Denhardt's reagent and 0.1 % SDS at 42 ° C or 1 to 2 hours in 6x SSC, 2xDenhardt's reagent and 0.1 % SDS at 68 °C. The denatured (dig- or radio-)labeled probe is then added directly to the prehybridization fluid and incubation is carried out for 16 to 24 hours at the appropriate temperature mentioned above. After incubation, the filters are then washed for 30 minutes at room temperature in 2x SSC, 0.1 % SDS, followed by 2 washes of 30 minutes each at 68 °C in 0.5 x SSC and 0.1 % SDS. An autoradiograph is established by exposing the filters for 24 to 48 hours to X-ray film (Kodak XAR-2 or equivalent) at -70 °C with an intensifying screen

"Insecticidal activity" of a protein, as used herein, means the capacity of a protein to kill insects when such protein is fed to insects, preferably by expression in a recombinant host such as a plant. "Insect-controlling amounts" of a protein, as used herein, refers to an amount of protein which is sufficient to limit damage on a plant by insects feeding on such plant to commercially acceptable levels, e.g. by killing the insects or by inhibiting the insect development or growth in such a manner that they provide less damage to a plant and plant yield is not significantly adversely affected.

In accordance with this invention, insects susceptible to the new Cry proteins of the invention are contacted with this protein in insect-controlling amounts, preferably insecticidal amounts.

"Cry protein" or "Cry protein of this invention", as used herein, refers to any one of the new proteins isolated in accordance with this invention and identified herein as Cry1Bf, Cry9Fa, or Cry1Jd protein. A Cry protein, as used herein, can be a protein in the full length size, also named a protoxin, or can be in a slightly or fully (e.g., N- and C-terminal truncation) truncated form as long as the insecticidal activity is retained, or can be a combination of different proteins or protein parts in a hybrid or fusion protein. A "Cry protoxin" refers to the full length crystal protein as it is encoded by the naturally-occurring *Bt* DNA sequence, a "Cry toxin" refers to an insecticidal fragment thereof, particularly the smallest toxic fragment thereof, typically in the molecular weight range of about 60 to about 80 kD as determined by SDS-PAGE electrophoresis. A "cry gene" or "cry DNA", as used herein, is a DNA sequence encoding a Cry protein in accordance with this invention, referring to any of the *cry1Bf*, *cry9Fa*, and *cry1Jd* DNA sequences defined above.

The "smallest toxic fragment" of a Cry protein, as used herein, is that fragment as can be obtained by trypsin or chymotrypsin digestion of the full length solubilized crystal protein that retains toxicity, or that toxic protein fragment encoded by DNA fragments of the Cry protein. This protein will mostly have a short N-terminal and a long C-terminal truncation compared to the protoxin. Although for recombinant expression, toxic fragments starting at or around original amino acid position 1 are a more preferred embodiment in accordance with this invention, it should be noted that besides a C-terminal truncation, some N-terminal amino acids can also be deleted while retaining the insecticidal character of the protein. The N-terminal end of the smallest toxic fragment is conveniently determined by N-terminal amino acid sequence determination of trypsin- or chymotrypsin-treated soluble crystal protein by techniques routinely available in the art.

Dna encoding the Cry proteins of this invention can be isolated in a conventional manner from the *E. coli* strains, deposited on November 25, 1999 at the BCCM-LMBP under accession numbers LMBP 3983, LMBP 3984, LMBP 3985 and LMBP 3986. The encoded Cry proteins can be used to prepare specific monoclonal or polyclonal antibodies in a conventional manner (Höfte et al., 1988). The toxin forms can be obtained by protease (e.g., trypsin) digestion of the Cry protoxins.

The DNA sequences encoding the Cry proteins can be isolated in a conventional manner from the respective strains or can be synthesized based on the encoded amino acid sequence.

The DNA sequences encoding the Cry proteins of the invention were identified by digesting total DNA from isolated *Bt* strains with restriction enzymes; size fractionating the DNA fragments, so produced, into DNA fractions of 5 to 10 Kb; ligating these fractions to cloning vectors; screening the *E. coli*, transformed with the cloning vectors, with a DNA probe that was constructed from a region of known *Bt* crystal protein genes or with a DNA probe based on specific PCR fragments generated from *Bt* DNA using primers corresponding to known *Bt* crystal protein genes.

Also, DNA sequences for use in this invention can be made synthetically. Indeed, because of the degeneracy of the genetic code, some amino acid codons can be replaced with others without changing the amino acid sequence of the protein. Furthermore, some amino acids can be substituted by other equivalent amino acids without significantly changing the insecticidal activity of the protein. Also, changes in amino acid sequence or composition in regions of the molecule, different from those responsible for binding and toxicity (e.g., pore formation) are less likely to cause a difference in insecticidal activity of the protein. Such equivalents of the gene include DNA sequences hybridizing to the DNA sequence of the Cry toxins or protoxins of SEQ ID. No. 2, 4, or 6 under stringent conditions and encoding a protein with the same insecticidal characteristics as the (pro)toxin of this invention, or DNA sequences encoding proteins with an amino acid sequence identity of at least 85 %, preferably at least 90 %, most preferably at least 95 %, with the protein toxin form (from the N-terminus to 2 amino acids beyond conserved sequence block 5 as defined in Schnepf et al., 1998) or with the protein protoxin form of the Cry1Bf, Cry9FA or Cry1Jd proteins of this invention, as determined using the GAP program of the Wisconsin package of GCG (Madison, Wisconsin, USA, version 10.0; GCG defaults were used within the GAP program; for the amino acid sequence comparisons, the blosum62 scoring matrix was used).

Of course, any other DNA sequence differing in its codon usage but encoding the same protein or a similar protein with substantially the same insecticidal activity, can be constructed, depending on the particular purpose. It has been described in prokaryotic and eucaryotic expression systems that changing the codon usage to that of the host cell is desired for gene

expression in foreign hosts (Bennetzen & Hall, 1982; Itakura, 1977). Furthermore, *Bt* crystal protein genes are known to have no bias towards eucaryotic codons, and to be very AT-rich (Adang et al., 1985, Schnepf et al., 1985). Codon usage tables are available in the literature (Wada et al., 1990; Murray et al., 1989) and in the major DNA sequence databanks (e.g. EMBL at Heidelberg, Germany). Accordingly, synthetic DNA sequences can be constructed so that the same or substantially the same proteins are produced. It is evident that several DNA sequences can be devised once the amino acid sequence of the Cry proteins of this invention is known. Such other DNA sequences include synthetic or semi-synthetic DNA sequences that have been changed in order to inactivate certain sites in the gene, e.g. by selectively inactivating certain cryptic regulatory or processing elements present in the native sequence as described in PCT publications WO 91/16432 and WO 93/09218, or by adapting the overall codon usage to that of a more related host organism, preferably that of the host organism in which expression is desired. When making such genes, the encoded amino acid sequence should be retained to the maximum extent possible, although truncations or minor replacements or additions of amino acids can be done as long as the toxicity of the protein is not negatively affected.

Small modifications to a DNA sequence such as described above can be routinely made by PCR-mediated mutagenesis (Ho et al., 1989, White et al., 1989).

With the term "substantially the same", when referring to a protein, is meant to include a protein that differs in some amino acids, or has some amino acids added (e.g. a fusion protein, see Vaeck et al., 1987) or deleted (e.g. N- or C-terminal truncation), as long as the protein has no major difference in its insecticidal activity.

The term "functional domain" of a Cry toxin as used herein means any part(s) or domain(s) of the toxin with a specific structure that can be transferred to another (Cry) protein for providing a new hybrid protein with at least one functional characteristic (e.g., the binding and/or toxicity characteristics) of the Cry toxin of the invention (Ge et al., 1991). Such parts can form an essential feature of the hybrid *Bt* protein with the binding and/or toxicity characteristics of the Cry protein of this invention. Such a hybrid protein can have an enlarged host range, an improved toxicity and/or can be used in a strategy to prevent insect resistance development (European Patent Publication ("EP") 408 403; Visser et al., 1993).

The 5 to 10 Kb fragments, prepared from total DNA of the *Bt* strains of the invention, can be ligated in suitable expression vectors and transformed in *E. coli*, and the clones can then be

screened by conventional colony immunoprobings methods (French et al., 1986) for expression of the toxin with monoclonal or polyclonal antibodies raised against the Cry proteins, or by hybridization with DNA probes.

Also, the 5 to 10 Kb fragments, prepared from total DNA of the *Bt* strains of the invention or fragments thereof cloned and/or subcloned in *E.coli*, can be ligated in suitable *Bt* shuttle vectors (Lereclus et al., 1992) and transformed in a crystal minus *Bt*-mutant. The clones are then screened for production of crystals (detected by microscopy) or crystal proteins (detected by SDS-PAGE).

The genes encoding the Cry proteins of this invention can be sequenced in a conventional manner (Maxam and Gilbert, 1980; Sanger, 1977) to obtain the DNA sequence. Sequence comparisons indicated that the genes are different from previously described genes encoding protoxins and toxins with activity against Lepidoptera (Höfte and Whiteley, 1989; Crickmore, et al., 1998); and the December 15, 1999 and October 16, 2000 updates on the *Bt* nomenclature website corresponding to the Crickmore et al. (1998) publication, found at:

http://epunix.biols.susx.ac.uk/Home/Neil_Crickmore/Bt/index.html

An insecticidally effective part of the DNA sequences, encoding an insecticidally effective portion of the newly identified Cry protein protoxin forms, can be made in a conventional manner after sequence analysis of the gene. In such fragments, it is preferred that at least the sequence up to the C-terminal end of conserved sequence block 5 of *Bt* proteins (Hofte & Whiteley, 1989; Schnepf et al., 1998), preferably up to two amino acids C-terminal of the conserved sequence block 5, is retained. The amino acid sequence of the Cry proteins can be determined from the DNA sequence of the isolated DNA sequences. By "an insecticidally effective part" of DNA sequences encoding the Cry protein, also referred to herein as "truncated gene" or "truncated DNA", is meant a DNA sequence encoding a polypeptide which has fewer amino acids than the Cry protein protoxin form but which is insecticidal to insects.

In order to express all or an insecticidally effective part of the DNA sequence encoding a Cry protein of this invention in *E. coli*, in other *Bt* strains and in plants, suitable restriction sites can be introduced, flanking the DNA sequence. This can be done by site-directed mutagenesis, using well-known procedures (Stanssens et al., 1989; White et al., 1989). In order to obtain improved expression in plants, the codon usage of the *cry* gene or insecticidally effective *cry* gene part of this invention can be modified to form an equivalent, modified or artificial gene or gene

part in accordance with PCT publications WO 91/16432 and WO 93/09218; EP 0 358 962 and EP 0 359 472, or the *Bt* genes or gene parts can be inserted in the chloroplast genome and expressed there using a chloroplast-active promoter (e.g., Mc Bride et al., 1995). For obtaining enhanced expression in monocot plants such as corn, a monocot intron also can be added to the chimeric gene, and the DNA sequence of the *cry* gene or its insecticidal part of this invention can be further changed in a translationally neutral manner, to modify possibly inhibiting DNA sequences present in the gene part by means of site-directed intron insertion and/or by introducing changes to the codon usage, e.g., adapting the codon usage to that most preferred by the specific plant (Murray et al., 1989) without changing significantly the encoded amino acid sequence.

Furthermore, the binding properties of the Cry proteins of the invention can be evaluated, using methods known in the art (Van Rie et al., 1990), to determine if the Cry proteins of the invention bind to sites on the insect midgut that are different from those recognized by other, known Cry or other *Bt* proteins. *Bt* toxins with different binding sites in relevant susceptible insects are very valuable to replace known *Bt* toxins to which insects may have developed resistance, or to use in combination with *Bt* toxins having a different mode of action to prevent or delay the development of insect resistance against *Bt* toxins, particularly when expressed in a plant. Because of the characteristics of the newly isolated *Bt* toxins, they are extremely useful for transforming plants, e.g. monocots such as corn or rice and vegetables such as *Brassica* species plants, to protect these plants from insect damage.

The insecticidally effective *cry* gene part or its equivalent, preferably the *cry* chimeric gene, encoding an insecticidally effective portion of the Cry protoxin, can be stably inserted in a conventional manner into the nuclear genome of a single plant cell, and the so-transformed plant cell can be used in a conventional manner to produce a transformed plant that is insect-resistant. In this regard, a disarmed Ti-plasmid, containing the insecticidally effective *cry* gene part, in *Agrobacterium tumefaciens* can be used to transform the plant cell, and thereafter, a transformed plant can be regenerated from the transformed plant cell using the procedures described, for example, in EP 0 116 718, EP 0 270 822, PCT publication WO 84/02913 and published European Patent application ("EP") 0 242 246 and in Gould et al. (1991). Preferred Ti-plasmid vectors each contain the insecticidally effective *cry* gene part between the border sequences, or at least located to the left of the right border sequence, of the T-DNA of the Ti-

plasmid. Of course, other types of vectors can be used to transform the plant cell, using procedures such as direct gene transfer (as described, for example in EP 0 233 247), pollen mediated transformation (as described, for example in EP 0 270 356, PCT publication WO 85/01856, and US Patent 4,684,611), plant RNA virus-mediated transformation (as described, for example in EP 0 067 553 and US Patent 4,407,956), liposome-mediated transformation (as described, for example in US Patent 4,536,475), and other methods such as the recently described methods for transforming certain lines of corn (Fromm et al., 1990; Gordon-Kamm et al., 1990) and rice (Shimamoto et al., 1989; Datta et al., 1990) and the recently described method for transforming monocots generally (PCT publication WO 92/09696).

The resulting transformed plant can be used in a conventional plant breeding scheme to produce more transformed plants with the same characteristics or to introduce the insecticidally effective *cry* gene part in other varieties of the same or related plant species. Seeds, which are obtained from the transformed plants, contain the insecticidally effective *cry* gene part as a stable genomic insert. Cells of the transformed plant can be cultured in a conventional manner to produce the insecticidally effective portion of the Cry protoxin, preferably the Cry toxin, which can be recovered for use in conventional insecticide compositions against Lepidoptera (US Patent 5,254,799). In accordance with this invention, plants or seeds of the invention can be used to obtain resistance to insects, e.g. by sowing or planting in a field wherein damaging insects usually occur, said seeds or plants. Methods for obtaining insect resistance and methods for obtaining improved yield or reduced insect damage are thus provided in accordance with the invention by planting or sowing in a field, preferably a field wherein damaging insects feeding on such plants usually occur or are expected to occur at levels which provide economic damage to the plants, the plants or seeds of the invention producing the Cry proteins of the invention.

The insecticidally effective *cry* gene part, preferably the truncated *cry* gene, is inserted in a plant cell genome so that the inserted gene is downstream (i.e., 3') of, and under the control of, a promoter which can direct the expression of the gene part in the plant cell. This is preferably accomplished by inserting the *cry* chimeric gene in the plant cell genome, particularly in the nuclear or chloroplast genome. Preferred promoters include: the strong constitutive 35S promoters (the "35S promoters") of the cauliflower mosaic virus (CaMV) of isolates CM 1841 (Gardner et al., 1981), CabbB-S (Franck et al., 1980) and CabbB-JI (Hull and Howell, 1987); promoters from the ubiquitin family (e.g., the maize ubiquitin promoter of Christensen

et al., 1992, see also Cornejo et al., 1993), the *gos2* promoter (de Pater et al., 1992), the *emu* promoter (Last et al., 1990), rice actin promoters such as the promoter described by Zhang et al. (1991); and the TR1' promoter and the TR2' promoter (the "TR1' promoter" and "TR2' promoter", respectively) which drive the expression of the 1' and 2' genes, respectively, of the T-DNA (Velten et al., 1984). Alternatively, a promoter can be utilized which is not constitutive but rather is specific for one or more tissues or organs of the plant (e.g., leaves and/or roots) whereby the inserted *cry* gene part is expressed only in cells of the specific tissue(s) or organ(s). For example, the insecticidally effective *cry* gene part could be selectively expressed in the leaves of a plant (e.g., corn, cotton) by placing the insecticidally effective gene part under the control of a light-inducible promoter such as the promoter of the ribulose-1,5-bisphosphate carboxylase small subunit gene of the plant itself or of another plant such as pea as disclosed in US Patent 5,254,799. Another alternative is to use a promoter whose expression is inducible (e.g., by temperature, wounding or chemical factors).

The insecticidally effective *cry* gene part is inserted in the plant genome so that the inserted gene part is upstream (i.e., 5') of suitable 3' end transcription regulation signals (i.e., transcript formation and polyadenylation signals). This is preferably accomplished by inserting the *cry* chimeric gene in the plant cell genome. Preferred polyadenylation and transcript formation signals include those of the octopine synthase gene (Gielen et al., 1984) and the T-DNA gene 7 (Velten and Schell, 1985), which act as 3'-untranslated DNA sequences in transformed plant cells.

The insecticidally effective *cry* gene part can optionally be inserted in the plant genome as a hybrid gene (US Patent 5,254,799; Vaeck et al., 1987) under the control of the same promoter as a selectable marker gene, such as the *neo* gene (EP 0 242 236) encoding kanamycin resistance, so that the plant expresses a fusion protein.

All or part of the *cry* gene, encoding an anti-lepidopteran protein, can also be used to transform other bacteria, such as a *B. thuringiensis* which has insecticidal activity against Lepidoptera or Coleoptera. Thereby, a transformed *Bt* strain can be produced which is useful for combatting a wide spectrum of lepidopteran and coleopteran insect pests or for combatting additional lepidopteran insect pests. Transformation of bacteria, such as bacteria of the genus *Agrobacterium*, *Bacillus* or *Escherichia*, with all or part of the *cry* gene of this invention, incorporated in a suitable cloning vehicle, can be carried out in a conventional manner, preferably

using conventional electroporation techniques as described in Mahillon et al. (1989) and in PCT Patent publication WO 90/06999.

Transformed *Bacillus* species strains containing the *cry* gene of this invention can be fermented by conventional methods (Dulmage, 1981; Bernhard and Utz, 1993) to provide high yields of cells. Under appropriate conditions which are well understood (Dulmage, 1981), these strains each sporulate to produce crystal proteins containing the Cry protoxin in high yields.

An insecticidal, particularly anti-lepidopteran, composition of this invention can be formulated in a conventional manner using the microorganisms transformed with the *cry* gene, or preferably their respective Cry proteins or the Cry protoxin, toxin or insecticidally effective protoxin portion as an active ingredient, together with suitable carriers, diluents, emulsifiers and/or dispersants (e.g., as described by Bernhard and Utz, 1993). This insecticide composition can be formulated as a wettable powder, pellets, granules or dust or as a liquid formulation with aqueous or non-aqueous solvents as a foam, gel, suspension, concentrate, etc.

A method for controlling insects, particularly Lepidoptera, in accordance with this invention can comprise applying (e.g., spraying), to a locus (area) to be protected, an insecticidal amount of the Cry proteins or host cells transformed with the *cry* gene of this invention. The locus to be protected can include, for example, the habitat of the insect pests or growing vegetation or an area where vegetation is to be grown.

To obtain the Cry protoxin or toxin, cells of the recombinant hosts expressing the Cry protein can be grown in a conventional manner on a suitable culture medium and then lysed using conventional means such as enzymatic degradation or detergents or the like. The protoxin can then be separated and purified by standard techniques such as chromatography, extraction, electrophoresis, or the like. The toxin can then be obtained by trypsin digestion of the protoxin.

The following Examples illustrate the invention, and are not provided to limit the invention or the protection sought. The sequence listing referred to in the Examples, the Claims and the Description is as follows:

Sequence Listing:

SEQ ID No. 1 - amino acid and DNA sequence of Cry1Bf protein and DNA

SEQ ID No. 2 - amino acid sequence of Cry1Bf protein.

SEQ ID No. 3 - amino acid and DNA sequence of Cry1Jd protein and DNA.
 SEQ ID No. 4 - amino acid sequence Cry1Jd protein.
 SEQ ID No. 5 - amino acid and DNA sequence of Cry9Fa protein and DNA.
 SEQ ID No. 6 - amino acid sequence of Cry9Fa protein.
 SEQ ID No. 7 - DNA sequence for primer Cry1B.fw.
 SEQ ID No. 8 - DNA sequence for primer B.R.
 SEQ ID No. 9 - DNA sequence for primer B.F.
 SEQ ID No. 10 - DNA sequence for primer JFW.
 SEQ ID No. 11 - DNA sequence for primer JRV.
 SEQ ID No. 12- DNA sequence for primer 9FW.
 SEQ ID No. 13 - DNA sequence for primer 9RV.

Unless otherwise stated in the Examples, all procedures for making and manipulating recombinant DNA are carried out by the standard procedures described in Sambrook et al., Molecular Cloning - A Laboratory Manual, Second Ed., Cold Spring Harbor Laboratory Press, NY (1989), and in Volumes 1 and 2 of Ausubel et al. (1994) Current Protocols in Molecular Biology, Current Protocols, USA. Standard materials and methods for plant molecular biology work are described in Plant Molecular Biology Labfax (1993) by R.R.D. Croy, jointly published by BIOS Scientific Publications Ltd (UK) and Blackwell Scientific Publications (UK). Procedures for PCR technology can be found in "PCR protocols: a guide to methods and applications", Edited by M.A. Innis, D.H. Gelfand, J.J. Sninsky and T.J. White (Academic Press, Inc., 1990).

EXAMPLES

Example 1: Characterization of the strains.

The BtS02072BG strain was isolated from a grain dust sample collected in Santo Tomas la Union, Ilocos, Philippines. The BtS02739C strain was isolated from a grain dust sample collected in Lucena City, South Tagalog, Philippines.

Each strain can be cultivated on conventional standard media, preferably T₃ medium (tryptone 3 g/l, tryptose 2 g/l, yeast extract 1.5 g/l, 5 mg MnCl₂, 0.05 M Na₂HPO₄·2H₂O, 0.05 M

NaH₂PO₄·H₂O, pH 6.8 and 1.5% agar), preferably at 28 °C. For long term storage, it is preferred to mix an equal volume of a spore-crystal suspension with an equal volume of 50% glycerol and store this at -70 °C or lyophilize a spore-crystal suspension. For sporulation, growth on T₃ medium is preferred for 72 hours at 28 °C, followed by storage at 4 °C. The crystal proteins produced by the strains during sporulation are packaged in crystals.

Example 2 : Insecticidal activity of the BtS02072BG and BtS02739C strains against selected lepidopteran insect species.

Toxicity assays were performed on neonate larvae of *Helicoverpa zea*, *Heliothis virescens*, *Ostrinia nubilalis*, *Spodoptera frugiperda* and *Sesamia nonagrioides* fed on an artificial diet layered with spore-crystal mixtures from either BtS02072BG or BtS02739C, at about 10⁹ spore-crystals per ml.

The artificial diet (Vanderzant, 1962) was dispensed in wells of Costar 24-well plates for tests on *H. zea*, *H. virescens* and *O. nubilalis*. 50 microliter of the spore-crystal mixture was applied on the surface of the diet and dried in a laminar air flow. For tests on *H. zea*, *H. virescens*, one larva was placed in each well and 20 larvae were used per sample. For tests on *O. nubilalis*, 2 larvae were placed in each well and 24 larvae were used per sample. The artificial diet was dispensed in wells of Costar 48-well plates for tests on *S. frugiperda* and *S. nonagrioides*. 25 microliter of the spore-crystal mixture was applied on the surface of the diet and dried in a laminar air flow. One larva was placed in each well and 18 larvae were used per sample. Dead and living larvae were counted on the seventh day. The percentage of dead larvae are shown in Table I below.

Table I: Percentage of dead larvae upon application of crystal-spore mixture to insects:

	BTS02072BG	BTS02739C
<i>H. zea</i>	70	15
<i>H. virescens</i>	85-50	80-60
<i>O. nubilalis</i>	92	72
<i>S. frugiperda</i>	6	Not tested
<i>S. nonagrioides</i>	100	Not tested

Example 3 : Characterization of new cry genes

The BtS02739C genes were detected by PCR using degenerate primers targeting conserved regions in known cry genes. The resulting amplification product was purified using the Wizard PCR preps (Promega) purification system and ligated into pGEM-T vector (Promega).

The ligation mixture was electroporated into *E. coli* JM101. A miniprep was made of at least 40 insert-containing transformants, and digests were performed with selected restriction enzymes. Following electrophoresis of the digested miniprep DNA, different DNA fragment patterns could be observed. For each pattern at least one colony was selected. An appropriate DNA prep was made in order to determine the sequence of the insert of the plasmid present in each selected colony. Alignment of the determined sequences of the amplification products with publicly available cry sequences demonstrates that strain BtS02739C contains a novel cry1J- type gene and a novel cry9- type gene.

The BtS02072BG gene was detected as follow. First, a PCR was performed using degenerate crystal protein gene primers on strain BtS02419J. The resulting amplification product was used as template in a secondary PCR using degenerate crystal protein primers

The resulting amplification product was purified using the Wizard PCR preps (Promega) purification system and ligated into pGEM-T vector (Promega). The ligation mixture was electroporated into XL1 Blue *E. coli*. A miniprep was made of at least 40 insert-containing transformants, and digests were performed with selected restriction enzymes. Following electrophoresis of the digested miniprep DNA, different DNA fragment patterns could be observed. For each pattern at least one colony was selected. An appropriate DNA prep was made in order to determine the sequence of the insert of the plasmid present in each selected colony.

From the cloned amplification products from strain BtS02419J, a sequence was found to be identical to the corresponding fragment of cry1Be1, except for one nucleotide difference. Next, primers were selected to evaluate the presence of a cry sequence similar to that of the sequenced cry gene fragment from BtS02419J in a number of Bt strains, one of them being strain BtS02072BG. These primers had the following sequence (5' to 3'):

Forward primer: cry1B.fw: CAG TCC AAA CGG GTA TAA AC

Reverse primer: B.R: CTG CTT CGA AGG TTG CAG TA

Alignment of the determined sequences from the amplification products with publicly available *cry* sequences demonstrates that strain BtS02072BG contains a novel *cry1B*-type gene.

Example 4 : Cloning and expression of the *cry* genes

In order to isolate the full length *cry1J*- type and *cry9*- type gene from BtS02739C, and the *cry1B*- type gene from BtS02072BG, total DNA from these strains was prepared and partially digested with *Sau*3A. The digested DNA was size fractionated on a sucrose gradient and fragments ranging from 5 Kb to 10 Kb were ligated to the *Bam*H1-digested and *Ts*AP (thermosensitive alkaline phosphatase)- treated cloning vector pUC19 (Yannisch-Perron et al, 1985). The ligation mixture was electroporated in *E. coli* XL1-Blue or *E. coli* JM109 cells. Transformants were plated on LB-triacillin plates containing Xgal and IPTG and white colonies were selected to be used in filter hybridization experiments. Recombinant *E.coli* clones containing the vector were then screened with the appropriate DIG labeled probes. These probes were prepared as follows. First, a PCR was performed using as template cells from a recombinant *E. coli* clone containing a plasmid harboring the particular *cry* gene fragment, previously amplified using appropriate primers as shown in Table II.

Table II: primers used to isolate novel *Bt* DNA sequences (Y= C or T, S= G or C):

strain	gene	primer	Length of amplified fragment	Primer sequence
2739C	<i>cry1J</i> -type	JFW	365 bp	GCA GCT AAT GCT ACC ACA TC
		JRV		GTG GCG GTA TGC TGA CTA AT
	<i>cry9</i> -type	9FW	576	GYT TTT ATT CGC CCG CCA CA
		9RV		CGA CAG TAG SAC CCA CTA CT
2072BG	<i>cry1B</i> -type	B.F	922	CAG CGT ATT AAG TCG ATG GA
		B.R		CTG CTT CGA AGG TTG CAG TA

The resulting amplification product was gel-purified and used as template in a secondary PCR reaction using DIG- labeled dNTPs. An appropriate amount of this amplification product was used in hybridization reactions.

Colony hybridization for strain BtS02739C was performed with a mixture of the *cry1J*- type probe and the *cry9*- type probe. Positive colonies were then hybridized with each probe separately. Colony hybridization for strain BtS02072BG was performed with the *cry1B*- type probe. Following identification of a positive colony containing a plasmid harboring the full length *cry* gene, the sequence of the *cry* gene was determined using the dye terminator labeling method and a Perkin Elmer ABI Prism-377 DNA sequencer for both strands. Upon DNA sequencing, the genes were termed as follows: the *cry1J*- type and *cry9*- type gene from BtS02739C were named *cry1Jd* and *cry9Fa*, respectively, and the *cry1B*- type gene from BtS02072BG was named *cry1Bf*. The genomic sequences of the isolated *cry1Jd*, *cry9Fa*, and *cry1Bf* genes, as well as the proteins they encode, are shown in the Sequence Listing included in this application. Comparison of the sequences with known Cry DNA or protein sequences showed that the sequences are novel and differ in a substantial number of nucleotides or amino acids from known *Bt* genes and proteins. Tables III-V provide an overview of the sequence identity with respect to the coding regions of the most similar genes and proteins (both protoxin as toxin forms) as determined using the GAP program of the Wisconsin package of GCG (Madison, Wisconsin, USA) version 10.0. GCG defaults were used within the GAP program. For nucleic acid sequence comparisons, the nwsgapdna scoring matrix was used, for amino acid sequence comparisons, the blosum62 scoring matrix. The toxin form, as used in Tables III-V, refers to the protein starting at the first amino acid and ending two amino acids beyond the last amino acid (usually a proline) of conserved sequence block 5, as defined in Schnepf et al. (1998). The protoxin form refers to the entire protein or coding region of the *Bt* protein/gene.

Table III: Sequence identities for *cry1Bf*/Cry1Bf:

DNA	<i>cry1Ba1</i>	<i>cry1Bb1</i>	<i>cry1Bc1</i>	<i>cry1Bd1</i>	<i>cry1Be1</i>
Protoxin	91.912	83.890	77.207	83.565	93.774
Toxin	86.562	74.922	74.922	75.342	89.220
Protein	Cry1Ba1	Cry1Bb1	Cry1Bc1	Cry1Bd1	Cry1Be1
Protoxin	89.869	80.193	75.795	80.933	92.170
Toxin	82.520	67.868	67.868	70.142	86.499

Table IV: Sequence identities for *cry9Fa*/Cry9Fa:

DNA	<i>cry9Aa1</i>	<i>cry9Ba1</i>	<i>cry9Ca1</i>	<i>cry9Da1</i>	<i>cry9Ea1</i>
Protoxin	71.592	78.212	76.614	81.197	84.043
Toxin	51.782	62.720	68.215	75.593	81.618
Protein	Cry9Aa1	Cry9Ba1	Cry9Ca1	Cry9Da1	Cry9Ea1
Protoxin	62.445	72.064	71.553	76.963	82.578
Toxin	35.828	52.167	59.133	68.372	78.858

Table V: Sequence identities for *cry1Jd*/Cry1Jd:

DNA	<i>cry1Ja1</i>	<i>cry1Jb1</i>	<i>cry1Jc1</i>
protoxin	83.233	83.176	86.323
toxin	79.526	81.162	88.143
protein	Cry1Ja1	Cry1Jb1	Cry1Jc1
protoxin	79.759	78.830	82.489
toxin	71.574	74.746	81.711

Genomic clones of the newly isolated genes have been deposited at the BCCM™-LMBP (Belgian Coordinated Collections of Microorganisms - Laboratorium voor Moleculaire Biologie-Plasmidencollectie, University of Gent, K.L. Ledeganckstraat 35, B-9000 Gent, Belgium) under the following accession numbers:

- LMBP 3983 for *E coli* JM109 containing plasmid pUC2739C/1Jd1 comprising the *cry1Jd* gene, deposited on November 25, 1999 (this gene can be isolated from this plasmid on an about 8.4 kb DNA fragment by digestion with XhoI and SmaI);
- LMBP 3984 for *E coli* JM109 containing plasmid pUC2739C/9Fa1 comprising the *cry9Fa* gene, deposited on November 25, 1999 (this gene can be isolated from this plasmid on an about 8 kb DNA fragment by digestion with SacI and PstI); and
- LMBP 3986 for *E coli* XL1Blue containing plasmid pUC2072BG/1Bf1 comprising the *cry1Bf* gene, deposited on November 25, 1999 (this gene can be isolated from this plasmid on an about 7 kb DNA fragment by digestion with SacI and Sall).

Example 5: Insecticidal activity of the cry genes:

The insert containing the *cry9Fa* gene was subcloned into a suitable shuttle vector and the resulting plasmid pSL2739C/9Fa1 was introduced by routine procedures into a crystal-minus *Bt* strain. The crystal protein produced by a sporulated culture of this recombinant *Bt* strain was tested on neonate larvae of *H. virescens* and *O. nubilalis* at a concentration of about 10^9 particles/ml. On *O. nubilalis* larvae, 100% mortality was observed, whereas 72% mortality was observed on *H. virescens* larvae, whereas after treatment with the crystal-minus control strain all larvae survived.

The insert containing the *cry1Bf* gene was subcloned into a suitable shuttle vector and the resulting plasmid pSL2072BG/1Bf was introduced by routine procedures into a crystal-minus *Bt* strain. The crystal protein produced by a sporulated culture of this recombinant *Bt* strain was tested on larvae of *Sesamia nonagrioides*, *Heliothis virescens*, *Helicoverpa zea* and *O. nubilalis* at different concentrations. Significant high mortality of the Cry1Bf toxin was observed on *H. virescens*, *Ostrinia nubilalis* and *Sesamia nonagrioides*, while lower toxicity was found on *Helicoverpa zea*. After treatment with the crystal-minus control strain all larvae survived.

The insert containing the *cry1Jd* gene was subcloned into a suitable shuttle vector and the resulting plasmid pGI2739C/1Jd was introduced by routine procedures into a crystal-minus *Bt* strain. The crystal protein produced by a sporulated culture of this recombinant *Bt* strain is tested on larvae of *Heliothis virescens* at different concentrations, and significant mortality of the Cry1Jd toxin was observed. After treatment with the crystal-minus control strain all larvae survived.

Example 6: production of the novel Cry proteins in transformed plants.

Chimeric genes encoding the truncated forms of the Cry1Bf, Cry1Jd, and Cry9Fa proteins are made as described in EP 0 193 259 and published PCT patent application WO 94/12264, using the CaMV 35S (Hull and Howell, 1987) and ubiquitin (Christensen et al., 1992)

promoters. Preferably, the codon usage of the open reading frame is adapted to that of the host plant so as to optimize expression efficiency, as described in published PCT patent application WO 94/12264.

Rice, cotton and corn cells are transformed with the resulting chimeric genes.

Corn cells are stably transformed by either *Agrobacterium*-mediated transformation (Ishida et al., 1996, and U.S. Patent No. 5,591,616) or by electroporation using wounded and enzyme-degraded embryogenic callus, as described in WO 92/09696 or US Patent 5,641,664 (incorporated herein by reference).

Cotton cells are stably transformed by *Agrobacterium*-mediated transformation (Umbeck et al., 1987, Bio/Technology 5, 263-266; US Patent 5,004,863, incorporated herein by reference).

Rice cells are stably transformed with the method described in published PCT patent application WO 92/09696.

Regenerated transformed corn, cotton and rice plants are selected by ELISA, Northern and Southern blot and insecticidal effect. Chimeric gene-containing progeny plants show improved resistance to insects compared to untransformed control plants with appropriate segregation of insect resistance and the transformed phenotype. Protein and RNA measurements show that increased insect resistance is linked with higher expression of the novel Cry protein in the plants.

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SEQUENCE LISTING

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<120> Insecticidal proteins from *Bacillus thuringiensis*.

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Gly Gly Asp Ile Ile Arg Thr Asn Val Asn Gly Ser Val Leu Ser Met	
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Gly Leu Asn Phe Asn Asn Thr Ser Leu Gln Arg Tyr Arg Val Arg Val	
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Ser Leu Thr Ser Gln Ser Phe Arg Phe Ala Glu Phe Pro Val Gly Ile	
595 600 605	
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ggt aga caa acg ttt cac ttt gat aaa att gaa ttc att cca att act	1920
Gly Arg Gln Thr Phe His Phe Asp Lys Ile Glu Phe Ile Pro Ile Thr	
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gca acc ttc gaa gca gaa tac gat tta gaa agg gcg caa gag gcg gtg	1968
Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Glu Ala Val	
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aat gct ctg ttt act aat acg aat cca aga aga ttg aaa aca gat gtg	2016
Asn Ala Leu Phe Thr Asn Thr Asn Pro Arg Arg Leu Lys Thr Asp Val	
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aca gat tat cat att gat caa gta tcc aat tta gtg gcg tgt tta tcg	2064
Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Ala Cys Leu Ser	
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Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Leu Glu Lys Val Lys	
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Tyr Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn	
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Phe Thr Ser Ile Asn Lys Gln Pro Asp Phe Ile Ser Thr Asn Glu Gln	
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Ser Asn Phe Thr Ser Ile His Glu Gln Ser Glu His Gly Trp Trp Gly	
740 745 750	
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785 790 795 800	
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Leu Arg Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile	
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tgc aga gat gga gaa aaa tgt gcg cat cat tcc cat cat ttc tct ttg	2640
Cys Arg Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu	
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Asp Ile Asp Ile Gly Cys Thr Asp Leu His Glu Asn Leu Gly Val Trp	
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Tyr Leu Ser Glu Leu Ser Val Ile Pro Gly Val Asn Ala Glu Ile Phe	
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Cys Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly	
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Tyr Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp	
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 Ser Tyr Ser His Arg Leu Ser His Ile Gly Leu Ile Ser Gln Ser Arg
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Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Ala Cys Leu Ser
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Cys Arg Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu
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Val	Lys	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Gln	930	935	940
Leu	Glu	Thr	Lys	Arg	Val	Tyr	Thr	Glu	Ala	Lys	Glu	Ala	Val	Asp	Ala	945	950	955
Leu	Phe	Val	Asp	Ser	Gln	Tyr	Asn	Arg	Leu	Gln	Ala	Asp	Thr	Asn	Ile	965	970	975
Gly	Met	Ile	His	Ala	Ala	Asp	Lys	Leu	Val	His	Arg	Ile	Arg	Glu	Ala	980	985	990
Tyr	Leu	Ser	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Glu	Ile	Phe	995	1000	1005
Glu	Glu	Leu	Glu	Gly	Arg	Ile	Ile	Thr	Ala	Ile	Ser	Leu	Tyr	Asp	Ala	1010	1015	1020
Arg	Asn	Val	Val	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ala	Cys	Trp	1025	1030	1035
Asn	Val	Lys	Gly	His	Val	Asp	Val	Gln	Gln	Ser	His	His	Arg	Ser	Val	1045	1050	1055
Leu	Val	Ile	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Ala	Val	Arg	Val	1060	1065	1070
Cys	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	1075	1080	1085
Tyr	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	1090	1095	1100
Glu	Leu	Lys	Phe	Lys	Asn	Cys	Glu	Glu	Glu	Glu	Val	Tyr	Pro	Thr	Asp	1105	1110	1115
Thr	Gly	Thr	Cys	Asn	Asp	Tyr	Thr	Ala	His	Gln	Gly	Thr	Ala	Val	Cys	1125	1130	1135
Asn	Ser	Arg	Asn	Ala	Gly	Tyr	Glu	Asp	Ala	Tyr	Glu	Val	Asp	Thr	Thr	1140	1145	1150
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 aag cct gag gaa gta ttt ttg gat ggg gag agg ata tta cct gat atc 96
 Lys Pro Glu Glu Val Phe Leu Asp Gly Glu Arg Ile Leu Pro Asp Ile
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 Asp Pro Leu Glu Val Ser Leu Ser Leu Leu Gln Phe Leu Leu Asn Asn
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 Phe Val Pro Gly Gly Gly Phe Ile Ser Gly Leu Ile Asp Lys Ile Trp
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 Gly Ala Leu Arg Pro Ser Glu Trp Glu Leu Phe Leu Ala Gln Ile Glu
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 Gln Leu Ile Asp Arg Arg Ile Glu Ala Thr Val Arg Ala Lys Ala Ile
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 Ala Glu Leu Glu Gly Leu Gly Arg Ser Tyr Gln Leu Tyr Gly Glu Ala
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Phe Lys Glu Trp Glu Lys Thr Pro Asp Asn Thr Ala Ala Arg Ser Arg	
115 120 125	
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Val Thr Glu Arg Phe Arg Ile Ile Asp Ala Gln Ile Glu Ala Asn Ile	
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Thr Tyr Lys Thr Glu Leu Glu Arg Leu Glu Phe Arg Ser Ile Ala Gln	
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Val Pro Pro His Arg Gly Tyr Ser His Leu Leu Ser His Val Thr Met	
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Glu Asn Tyr Val Thr Leu Pro Gly Thr Phe Asp Ala Cys Tyr Pro Thr	
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Cys Ser Cys Arg Asp Gly Glu Lys Cys Ala His His Ser His His Phe	
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Cys Trp Asn Val Lys Gly His Val Asp Ile Lys Gln Asn Gly His Arg	
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 Val Thr Glu Arg Phe Arg Ile Ile Asp Ala Gln Ile Glu Ala Asn Ile
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 Pro Ser Phe Arg Val Ser Gly Phe Glu Val Pro Leu Leu Ser Val Tyr
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 Phe Gly Glu Arg Trp Gly Leu Ser Thr Thr Asn Val Asn Asp Ile Tyr
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 Asn Arg Gln Val Lys Arg Ile His Glu Tyr Ser Asp His Cys Val Asp
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 Trp Arg Ile Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val Leu
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 Ala Ser His Leu Ser Gly Gly Ser Val Ile Lys Gly Pro Gly His Thr
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Pro Asn Ala Val Phe Gln Asn Met His Tyr Lys Asp Tyr Leu Gln Thr
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Tyr Asp Gly Asp Tyr Thr Gly Ser Phe Ile Asn Pro Asn Leu Ser Ile
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Asp Gln Ala Ala Asn Leu Val Ser Cys Leu Ser Asp Glu Gln Tyr Ala	
690 695 700	

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Lys	Asn	Val	Leu	Asp	Asn	Leu	Val	Ser	Asp	Thr	Tyr	Pro	Asp	Asp	Ser	
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Cys	Ser	Gly	Ile	Asn	Arg	Cys	Glu	Glu	Gln	Gln	Met	Val	Asn	Ala	Gln	
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Leu	Glu	Thr	Glu	His	His	His	Pro	Met	Asp	Cys	Cys	Glu	Ala	Ala	Gln	
			850			855						860				
aca	cat	gag	ttt	tct	tcc	tat	att	gat	aca	ggg	gat	tta	aat	tcg	act	2640
Thr	His	Glu	Phe	Ser	Ser	Tyr	Ile	Asp	Thr	Gly	Asp	Leu	Asn	Ser	Thr	
865						870				875					880	
gta	gac	cag	gga	atc	tgg	gtg	atc	ttt	aaa	gtt	cga	aca	aca	gat	gg	2688
Val	Asp	Gln	Gly	Ile	Trp	Val	Ile	Phe	Lys	Val	Arg	Thr	Thr	Asp	Gly	
				885					890						895	
tat	gcg	acg	cta	gga	aat	ctt	gaa	ttg	gta	gag	gtc	gga	ccg	tta	ttg	2736
Tyr	Ala	Thr	Leu	Gly	Asn	Leu	Glu	Leu	Val	Glu	Val	Gly	Pro	Leu	Leu	
			900					905							910	

ggt gaa cct cta gaa cgt gaa caa aga gaa aat gcg aaa tgg aat gca	2784
Gly Glu Pro Leu Glu Arg Glu Gln Arg Glu Asn Ala Lys Trp Asn Ala	
915 920 925	
gag tta gga aga aaa cgt gca gaa aca gat cgc gtg tat caa gat gcc	2832
Glu Leu Gly Arg Lys Arg Ala Glu Thr Asp Arg Val Tyr Gln Asp Ala	
930 935 940	
aaa caa tcc atc aat cat tta ttt gtg gat tat caa gat caa caa tta	2880
Lys Gln Ser Ile Asn His Leu Phe Val Asp Tyr Gln Asp Gln Gln Leu	
945 950 955 960	
aat cca caa ata ggg atg gca gat att atg gac gct caa aat ctt gtc	2928
Asn Pro Gln Ile Gly Met Ala Asp Ile Met Asp Ala Gln Asn Leu Val	
965 970 975	
gca tca att tca gat gta tat agc gat gca gta ctg caa atc cct gga	2976
Ala Ser Ile Ser Asp Val Tyr Ser Asp Ala Val Leu Gln Ile Pro Gly	
980 985 990	
att aac tat gag att tac aca gag ctg tcc aat cgc tta caa caa gca	3024
Ile Asn Tyr Glu Ile Tyr Thr Glu Leu Ser Asn Arg Leu Gln Gln Ala	
995 1000 1005	
tcg tat ctg tat acg tct cga aat gcg gtg caa aat ggg gac ttt aac	3072
Ser Tyr Leu Tyr Thr Ser Arg Asn Ala Val Gln Asn Gly Asp Phe Asn	
1010 1015 1020	
aac ggg cta gat agc tgg aat gca aca gcg ggt gca tcg gta caa cag	3120
Asn Gly Leu Asp Ser Trp Asn Ala Thr Ala Gly Ala Ser Val Gln Gln	
1025 1030 1035 1040	
gat ggc aat acg cat ttc tta gtt ctt tct cat tgg gat gca caa gtt	3168
Asp Gly Asn Thr His Phe Leu Val Leu Ser His Trp Asp Ala Gln Val	
1045 1050 1055	
tcc caa caa ttt aga gtg cag ccg aat tgt aaa tat gta tta cgt gta	3216
Ser Gln Gln Phe Arg Val Gln Pro Asn Cys Lys Tyr Val Leu Arg Val	
1060 1065 1070	
aca gca gag aaa gta ggc ggc gga gac gga tac gtg act atc cgg gat	3264
Thr Ala Glu Lys Val Gly Gly Gly Asp Gly Tyr Val Thr Ile Arg Asp	
1075 1080 1085	
ggt gct cat cat aca gaa acg ctt aca ttt aat gca tgt gat tat gat	3312
Gly Ala His His Thr Glu Thr Leu Thr Phe Asn Ala Cys Asp Tyr Asp	
1090 1095 1100	
ata aat ggc acg tac gtg act gat aat acg tat cta aca aaa gaa gtg	3360
Ile Asn Gly Thr Tyr Val Thr Asp Asn Thr Tyr Leu Thr Lys Glu Val	
1105 1110 1115 1120	

ata ttc tat tca cat aca gaa cac atg tgg gta gag gta aat gaa aca 3408
 Ile Phe Tyr Ser His Thr Glu His Met Trp Val Glu Val Asn Glu Thr
 1125 1130 1135

 gaa ggt gca ttt cat ata gat agt att gaa ttc gtt gaa aca gaa aag 3456
 Glu Gly Ala Phe His Ile Asp Ser Ile Glu Phe Val Glu Thr Glu Lys
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 taa 3459

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 Cys Gly Cys Ala Ser Asp Asp Val Val Gln Tyr Pro Leu Ala Arg Asp
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 Pro Asn Ala Val Phe Gln Asn Met His Tyr Lys Asp Tyr Leu Gln Thr
 35 40 45

 Tyr Asp Gly Asp Tyr Thr Gly Ser Phe Ile Asn Pro Asn Leu Ser Ile
 50 55 60

 Asn Pro Arg Asp Val Leu Gln Thr Gly Ile Asn Ile Val Gly Arg Leu
 65 70 75 80

 Leu Gly Phe Leu Gly Val Pro Phe Ala Gly Gln Leu Val Thr Phe Tyr
 85 90 95

 Thr Phe Leu Leu Asn Gln Leu Trp Pro Thr Asn Asp Asn Ala Val Trp
 100 105 110

 Glu Ala Phe Met Ala Gln Ile Glu Glu Leu Ile Asn Gln Arg Ile Ser
 115 120 125

 Glu Ala Val Val Gly Thr Ala Ala Asp His Leu Thr Gly Leu His Asp
 130 135 140

 Asn Tyr Glu Leu Tyr Val Glu Ala Leu Glu Glu Trp Leu Glu Arg Pro
 145 150 155 160

 Asn Ala Ala Arg Thr Asn Leu Leu Phe Asn Arg Phe Thr Thr Leu Asp
 165 170 175

 Ser Leu Phe Thr Gln Phe Met Pro Ser Phe Gly Thr Gly Pro Gly Ser
 180 185 190

Gln Asn Tyr Ala Val Pro Leu Leu Thr Val Tyr Ala Gln Ala Ala Asn
 195 200 205
 Leu His Leu Leu Leu Lys Asp Ala Glu Ile Tyr Gly Ala Arg Trp
 210 215 220
 Gly Leu Asn Gln Asn Gln Ile Asn Ser Phe His Thr Arg Gln Gln Glu
 225 230 235 240
 Arg Thr Gln Tyr Tyr Thr Asn His Cys Val Thr Thr Tyr Asn Thr Gly
 245 250 255
 Leu Asp Arg Leu Arg Gly Thr Asn Thr Glu Ser Trp Leu Asn Tyr His
 260 265 270
 Arg Phe Arg Arg Glu Met Thr Leu Met Ala Met Asp Leu Val Ala Leu
 275 280 285
 Phe Pro Tyr Tyr Asn Val Arg Gln Tyr Pro Asn Gly Ala Asn Pro Gln
 290 295 300
 Leu Thr Arg Glu Ile Tyr Thr Asp Pro Ile Val Tyr Asn Pro Pro Ala
 305 310 315 320
 Asn Gln Gly Ile Cys Arg Arg Trp Gly Asn Asn Pro Tyr Asn Thr Phe
 325 330 335
 Ser Glu Leu Glu Asn Ala Phe Ile Arg Pro Pro His Leu Phe Asp Arg
 340 345 350
 Leu Asn Arg Leu Thr Ile Ser Arg Asn Arg Tyr Thr Ala Pro Thr Thr
 355 360 365
 Asn Ser Tyr Leu Asp Tyr Trp Ser Gly His Thr Leu Gln Ser Gln Tyr
 370 375 380
 Ala Asn Asn Pro Thr Thr Tyr Glu Thr Ser Tyr Gly Gln Ile Thr Ser
 385 390 395 400
 Asn Thr Arg Leu Phe Asn Thr Thr Asn Gly Ala Asn Ala Ile Asp Ser
 405 410 415
 Arg Ala Arg Asn Phe Gly Asn Leu Tyr Ala Asn Leu Tyr Gly Val Ser
 420 425 430
 Tyr Leu Asn Ile Phe Pro Thr Gly Val Met Ser Glu Ile Thr Ser Ala
 435 440 445
 Pro Asn Thr Cys Trp Gln Asp Leu Thr Thr Thr Glu Glu Leu Pro Leu
 450 455 460

Val Asn Asn Asn Phe Asn Leu Leu Ser His Val Thr Phe Leu Arg Phe
465 470 475 480
Asn Thr Thr Gln Gly Gly Pro Leu Ala Thr Val Gly Phe Val Pro Thr
485 490 495
Tyr Val Trp Thr Arg Gln Asp Val Asp Phe Asn Asn Ile Ile Thr Pro
500 505 510
Asn Arg Ile Thr Gln Ile Pro Val Val Lys Ala Tyr Glu Leu Ser Ser
515 520 525
Gly Ala Thr Val Val Lys Gly Pro Gly Phe Thr Gly Gly Asp Val Ile
530 535 540
Arg Arg Thr Asn Thr Gly Gly Phe Gly Ala Ile Arg Val Ser Val Thr
545 550 555 560
Gly Pro Leu Thr Gln Arg Tyr Arg Ile Arg Phe Arg Tyr Ala Ser Thr
565 570 575
Ile Asp Phe Asp Phe Phe Val Thr Arg Gly Gly Thr Thr Ile Asn Asn
580 585 590
Phe Arg Phe Thr Arg Thr Met Asn Arg Gly Gln Glu Ser Arg Tyr Glu
595 600 605
Ser Tyr Arg Thr Val Glu Phe Thr Thr Pro Phe Asn Phe Thr Gln Ser
610 615 620
Gln Asp Ile Ile Arg Thr Ser Ile Gln Gly Leu Ser Gly Asn Gly Glu
625 630 635 640
Val Tyr Leu Asp Arg Ile Glu Ile Ile Pro Val Asn Pro Thr Arg Glu
645 650 655
Ala Glu Glu Asp Leu Glu Ala Ala Lys Lys Ala Val Ala Ser Leu Phe
660 665 670
Thr Arg Thr Arg Asp Gly Leu Gln Val Asn Val Thr Asp Tyr Gln Val
675 680 685
Asp Gln Ala Ala Asn Leu Val Ser Cys Leu Ser Asp Glu Gln Tyr Ala
690 695 700
His Asp Lys Lys Met Leu Leu Glu Ala Val Arg Ala Ala Lys Arg Leu
705 710 715 720
Ser Arg Glu Arg Asn Leu Leu Gln Asp Pro Asp Phe Asn Thr Ile Asn
725 730 735
Ser Thr Glu Glu Asn Gly Trp Lys Ala Ser Asn Gly Val Thr Ile Ser
740 745 750

Glu Gly Gly Pro Phe Tyr Lys Gly Arg Ala Leu Gln Leu Ala Ser Ala
 755 760 765
 Arg Glu Asn Tyr Pro Thr Tyr Ile Tyr Gln Lys Val Asp Ala Ser Glu
 770 775 780
 Leu Lys Pro Tyr Thr Arg Tyr Arg Leu Asp Gly Phe Val Lys Ser Ser
 785 790 795 800
 Gln Asp Leu Glu Ile Asp Leu Ile His His His Lys Val His Leu Val
 805 810 815
 Lys Asn Val Leu Asp Asn Leu Val Ser Asp Thr Tyr Pro Asp Asp Ser
 820 825 830
 Cys Ser Gly Ile Asn Arg Cys Glu Glu Gln Gln Met Val Asn Ala Gln
 835 840 845
 Leu Glu Thr Glu His His His Pro Met Asp Cys Cys Glu Ala Ala Gln
 850 855 860
 Thr His Glu Phe Ser Ser Tyr Ile Asp Thr Gly Asp Leu Asn Ser Thr
 865 870 875 880
 Val Asp Gln Gly Ile Trp Val Ile Phe Lys Val Arg Thr Thr Asp Gly
 885 890 895
 Tyr Ala Thr Leu Gly Asn Leu Glu Leu Val Glu Val Gly Pro Leu Leu
 900 905 910
 Gly Glu Pro Leu Glu Arg Glu Gln Arg Glu Asn Ala Lys Trp Asn Ala
 915 920 925
 Glu Leu Gly Arg Lys Arg Ala Glu Thr Asp Arg Val Tyr Gln Asp Ala
 930 935 940
 Lys Gln Ser Ile Asn His Leu Phe Val Asp Tyr Gln Asp Gln Gln Leu
 945 950 955 960
 Asn Pro Gln Ile Gly Met Ala Asp Ile Met Asp Ala Gln Asn Leu Val
 965 970 975
 Ala Ser Ile Ser Asp Val Tyr Ser Asp Ala Val Leu Gln Ile Pro Gly
 980 985 990
 Ile Asn Tyr Glu Ile Tyr Thr Glu Leu Ser Asn Arg Leu Gln Gln Ala
 995 1000 1005
 Ser Tyr Leu Tyr Thr Ser Arg Asn Ala Val Gln Asn Gly Asp Phe Asn
 1010 1015 1020

Asn Gly Leu Asp Ser Trp Asn Ala Thr Ala Gly Ala Ser Val Gln Gln
 025 1030 1035 1040

Asp Gly Asn Thr His Phe Leu Val Leu Ser His Trp Asp Ala Gln Val
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Ser Gln Gln Phe Arg Val Gln Pro Asn Cys Lys Tyr Val Leu Arg Val
 1060 1065 1070

Thr Ala Glu Lys Val Gly Gly Gly Asp Gly Tyr Val Thr Ile Arg Asp
 1075 1080 1085

Gly Ala His His Thr Glu Thr Leu Thr Phe Asn Ala Cys Asp Tyr Asp
 1090 1095 1100

Ile Asn Gly Thr Tyr Val Thr Asp Asn Thr Tyr Leu Thr Lys Glu Val
 1105 1110 1115 1120

Ile Phe Tyr Ser His Thr Glu His Met Trp Val Glu Val Asn Glu Thr
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Glu Gly Ala Phe His Ile Asp Ser Ile Glu Phe Val Glu Thr Glu Lys
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 cry1B.fw

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<210> 8

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gcagctaata ctaccacatc

20

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gyttttattc gcccgccaca

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cgacagtags acccactact

20